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(21) International Application Number: PCT/US97/02562 (22) International Filing Date: 26 February 1997 (26.02.97) (30) Priority Data: 08/609,806 1 March 1996 (01.03.96) US (71) Applicant: SCHERING CORPORATION [US/US]; 2000 Galloping Hill Road, Kenilworth, NJ 07033 (US). (72) Inventor: HESSE, Richard, A.; 1229 South 164th Avenue, Omaha, NE 68130 (US). (74) Agents: MAITNER, John, J. et al.; Schering-Plough Corporation, Patent Dept. K-6-1 1990, 2000 Galloping Hill Road, Kenilworth, NJ 07033-0530 (US).	(81) Designated States: AL, AM, AU, AZ, BA, BB, BG, BR, BY, CA, CN, CZ, EE, GE, HU, IL, IS, JP, KG, KR, KZ, LC, LK, LR, LT, LV, MD, MG, MK, MN, MX, NO, NZ, PL, RO, RU, SG, SI, SK, TJ, TM, TR, TT, UA, UZ, VN, YU, ARIPO patent (GH, KE, LS, MW, SD, SZ, UG), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG).	
Published <i>With international search report.</i> <i>Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.</i>		(54) Title: PORCINE REPRODUCTIVE AND RESPIRATORY SYNDROME VACCINE (57) Abstract The invention discloses a vaccine and methods for the treatment of porcine reproductive and respiratory syndrome. The vaccine is derived from the viral agent NEB-1-P94, deposited at the American Type Culture Collection under accession number VR-2525. Further, the invention discloses a vaccine virus with phenotypic characteristics which can be distinguished from wild type PRRS virus.

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PORCINE REPRODUCTIVE AND RESPIRATORY SYNDROME VACCINE

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BACKGROUND OF THE INVENTION

The present invention relates to a vaccine for the treatment of porcine reproductive and respiratory syndrome (PRRS).

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In 1987, the swine-producing industry in the United States experienced an unknown infectious disease which had a serious economic impact on the swine industry. The disease syndrome was reported in Europe including Germany, Belgium, the Netherlands, Spain and England.

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The disease is characterized by reproductive failure, respiratory disease and various clinical signs including loss of appetite, fever, dyspnea, and mild neurologic signs. A major component of the syndrome is reproductive failure which manifests itself as premature births, late term abortions, pigs born weak, stillbirths, mummified fetuses, decreased farrowing rates, and delayed return of estrus. Clinical signs of respiratory disease are most pronounced in pigs under 3-weeks-of-age but are reported to occur in pigs at all stages of production. Affected piglets grow slowly, have roughened hair coats, respiratory distress ("thumping"), and increased mortality.

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The disease syndrome has been referred to by many different terms including mystery swine disease (MSD), porcine epidemic abortion and respiratory syndrome (PEARS), swine infertility and respiratory syndrome (SIRS). The name now commonly used is porcine reproductive and respiratory syndrome (PRRS); this term will be employed throughout this patent application.

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It is an object of the invention to provide a vaccine which protects a pig against clinical disease caused by PRRS. Another object is to provide a vaccine which, when administered to a

breeding swine herd, will reduce the presence of PRRS in their population.

SUMMARY OF THE INVENTION

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An object of the present invention is to provide a novel vaccine which protects a pig against clinical disease caused by porcine reproductive and respiratory syndrome (PRRS) virus.

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A further object of the present invention is to provide a vaccine which protects a pig against the strain NEB-1 of PRRS virus.

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It is a further object of the present invention to provide a method of protecting a pig against clinical disease caused by a porcine reproductive and respiratory disease virus.

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It is a further object of this invention to provide a vaccine virus with phenotypic characteristics which can be distinguished from wild type PRRS virus.

DETAILED DESCRIPTION OF THE INVENTION

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The invention provides a composition of matter comprising an attenuated Porcine Respiratory and Reproductive Syndrome (PRRS) Virus which has been modified by laboratory manipulation for use in vaccination. Further the composition has phenotypic properties which allow its use for diagnostic purposes to distinguish between swine that have been naturally-infected with PRRS virus versus animals that have only been exposed to the vaccine strain. The PRRS virus isolate NEB-1-P94 has been deposited with American Type Culture Collection (Accession No. VR-2525).

35

A virulent isolate of PRRS virus was obtained from tissue samples from a dead pig presented to the University of Nebraska Diagnostic Laboratory. A tissue homogenate from the dead pig was inoculated onto primary swine alveolar macrophages and the presence of virus detected by cytopathic effects on inoculated

but not control cultures. The isolated virus (designated NEB-1) was subsequently characterized as a PRRS virus based on physical properties (ether and chloroform sensitivity, buoyant density, and lack of hemagglutinating activity), reactivity with specific
5 antibodies, and genetic analysis. Inoculation of the virus into nursing piglets resulted in a respiratory disease characterized by high fever, altered respiration and lung pathology consistent with viral interstitial pneumonia. Additionally, inoculation of the virus into pregnant sows resulted in reproductive disease
10 characterized by mummification of fetuses, stillborn piglets, and piglets born weak that subsequently died. The respiratory and reproductive disease caused by this virus was typical of the syndrome reported for PRRS virus.

15 The NEB-1 virus was attenuated by serial passage in tissue culture. The virus was initially passed by inoculation of primary swine alveolar macrophage (SAM) cultures (for the first two passages) and then by serial passage on MA104 cells (available from Microbiological Associates, Inc., Rockville, MD)
20 for a total of 94 passages. During this process, virus clones were isolated by plaque purification and characterized for phenotypic properties. The vaccine clone, designated NEB-1-P94, was selected for impaired growth on swine alveolar macrophages, lack of reactivity with the PRRS-specific monoclonal antibody
25 SDOW17 (ATCC HB10997), and lack of disease induction in piglets and pregnant sows. The NEB-1-P94 was expanded on MA104 cells and frozen as a master seed virus, also designated PRRS-MSV-94-1, for use in vaccine development studies.

30 Vaccine is prepared using MA104 cells as the substrate (however alternate cell lines that support the growth of PRRS virus such as MARC 145 [available from Dr. Wang, Agriculture Research Station, Clay Center NE] cells can also be used). MA104 cells are grown to confluency in suitable tissue culture vessels,
35 e.g. 850 cm² roller bottles, using Eagle's minimum essential media (EMEM) containing 5 to 10% bovine serum, 30 mM HEPES (N-2-hydroxyethylpiperazine-N'-2-ethanesulfonic acid, 2 mM L-

glutamine, and antibiotics (such as 30 µg/ml gentamicin). Alternate tissue culture media that can support the growth of MA104 cells such as Dulbecco's modified essential media [DMEM], Medium 199, or others can also be used. Confluent monolayers of MA104 cells are inoculated with NEB-1-P94 virus at a multiplicity of infection (MOI) in the range of 1:5 to 1:1000, and, preferably in the range of 1:10. Following incubation for three to five days at 37°C, culture supernatant fluids are harvested by decanting.

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Virus fluids are titered by making serial dilutions in EMEM supplemented as above and inoculation of 0.2 ml per well into at least four replicate wells of confluent MA104 or MARC 145 cells in a 96-well tissue culture plate. Cultures are incubated for five days at 37°C, 3-5% CO₂ in a humidified chamber and observed for cytopathic effects. Titers (50% endpoints) are calculated according to the methods of Spearman and Karber (Methods in Virology, Volume IV, K. Maramorosch and H. Koprowski (Eds). Academic Press, New York, 1977). Cells may be fixed with 80% acetone and tested for lack of reactivity with SDOW17 and positive reactivity with V017 or EP147 (available from Dr. E. Nelson, South Dakota State University, Brookings, SD) (expected positive result) monoclonal antibodies to confirm phenotypic identity of the virus.

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For the preparation of a killed vaccine, virus fluids are incubated with a chemical inactivation agent. Examples of inactivation agents include formaldehyde, glutaraldehyde, binary ethyleneimine, or beta-propiolactone. Virus fluids are then stored at 4°C until formulated into vaccine. Vaccine is prepared by mixing virus fluids (containing 10⁶ to 10⁹ TCID₅₀ of virus; based on preinactivation titers) with a physiologically acceptable diluent (such as EMEM, Hank's Balanced Salt Solution, Phosphate Buffered Saline) and an immune-stimulating adjuvant (such as mineral oil, vegetable oil, aluminum hydroxide, saponin, non-ionic detergents, squalene, block co-polymers or other compounds known in the art, used alone or in combination). A vaccine dose is typically between 1 and 5 ml.

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For a live vaccine formulation, virus fluids are stored frozen at -50°C or colder until use. Virus fluids within the range of $10^{4.0}$ and $10^{7.0}$ TCID₅₀/dose and preferably containing $10^{6.0}$ TCID₅₀/dose are diluted with a physiologically suitable diluent (such as EMEM, Hank's Balanced Salt Solution, Phosphate Buffered Saline) and a physiologically suitable mixture of compounds designed to stabilize the virus. Compounds known in the art that can be used alone or in combination to stabilize viruses include sucrose, lactose, N-Z amine, glutathione, neopeptone, gelatin, dextran and tryptone. Vaccine is stored frozen (-50°C or colder) or lyophilized with storage at 4°C until use. The vaccine typically has a dose size range of 1 to 5 ml, and preferably 2 ml.

For prophylaxis against PRRS-induced disease, the vaccine is administered to the pig orally, intranasally or parenterally. Examples of parenteral routes of administration include intradermal, intramuscular, intravenous, intraperitoneal and subcutaneous routes of administration.

When administered as a solution, the present vaccine may be prepared in the form of an aqueous solution, a syrup, an elixir, or a tincture. Such formulations are known in the art, and are prepared by dissolution of the antigen and other appropriate additives in the appropriate solvent systems. Such solvents include water, saline, ethanol, ethylene glycol, glycerol, A1 fluid, etc. Suitable additives known in the art include certified dyes, flavors, sweeteners, and antimicrobial preservatives, such as thimerosal (sodium ethylmercuri-thiosalicylate). Such solutions may be stabilized, for example, by addition of partially hydrolyzed gelatin, sorbitol, or cell culture medium, and may be buffered by methods known in the art, using reagents known in the art, such as sodium hydrogen phosphate, sodium dihydrogen phosphate, potassium hydrogen phosphate and/or potassium dihydrogen phosphate.

Liquid formulations may also include suspensions and emulsions. The preparation of suspensions, for example using a

colloid mill, and emulsions, for example using a homogenizer, is known in the art.

5 Parenteral dosage forms, designed for injection into body fluid systems, require proper isotonicity and pH buffering to the corresponding levels of porcine body fluids. Parenteral formulations must also be sterilized prior to use.

10 Isotonicity can be adjusted with sodium chloride and other salts as needed. Other solvents, such as ethanol or propylene glycol, can be used to increase solubility of ingredients of the composition and stability of the solution. Further additives which can be used in the present formulation include dextrose, conventional antioxidants and conventional chelating agents, such
15 as ethylenediamine tetraacetic acid (EDTA).

A booster vaccination may be administered two to four weeks after the initial immunization. For the prevention of reproductive disease, the vaccination regimen is typically
20 performed up to 6 weeks prior to and 1 week after breeding. For the prevention of respiratory disease in piglets, vaccination may be given as early as 3 weeks of age. The response to vaccination can be monitored by measuring antibody titer directed against PRRS virus using enzyme-linked immunosorbent assay (ELISA), serum
25 neutralization assay, indirect immunofluorescence, or Western blot.

The vaccine strain has phenotypic properties that can be used for diagnosis in swine of wild type PRRS infection versus
30 exposure to only the vaccine strain. Animals exposed to field strains of PRRS virus may be distinguished from animals exposed only to the NEB-1-P94 vaccine strain by measurement of the antibody response to the epitope recognized by monoclonal antibody (MAb) SDOW17. The presence of antibodies reactive with
35 the SDOW17 epitope is indicative of wild type virus exposure,

Measurement of antibodies to the SDOW17 epitope can be accomplished using a competitive ELISA. Plates (96-well) are coated with the NEB-1 PRRS virus (or other PRRS viruses

expressing the SDOW17 epitope). Plates are then incubated with pig serum from test animals and enzyme-labelled (for example conjugated to horseradish peroxidase) SDOW17 monoclonal antibody. The ability of pig sera to recognize the SDOW17 epitope is

5 measured by the inhibition of enzyme-linked SDOW17 MAb binding to the plate as detected by lack of enzyme substrate color conversion. Alternatively, a direct ELISA may be used. The amino acid sequence comprising the SDOW17 epitope can be prepared as a synthetic peptide or by recombinant DNA expression methods

10 in a suitable vector system such as *E. coli*. Plates coated with the SDOW17 antigen are incubated with pig serum. Binding of swine antibodies to the SDOW17 antigen is detected by incubation with enzyme-conjugated anti-swine immunoglobulin antisera followed by incubation with enzyme substrate and detection of a

15 color change.

The following examples describe in detail the invention. It will be apparent to those skilled in the art that modification of materials and methods may be practiced without departing from the

20 purpose and intent of this disclosure.

EXAMPLE 1

Phenotypic Characterization of NEB-1-P94 for Growth on
Alveolar Macrophages

The NEB-1-P94 vaccine strain virus at five passages from the master seed was characterized for growth on MA104, MARC 145, and swine alveolar macrophages. Swine alveolar macrophages (SAM) were obtained by bronchi-alveolar lavage with saline followed by centrifugation to pellet the cells. Macrophages were resuspended in EMEM with 10% fetal bovine serum and 50 µg/ml gentamicin and plated at approximately 7×10^4 cells per well of 96-well tissue culture plates. MA104 cells and MARC 145 cells were plated in 96-well tissue culture plates in media (EMEM containing 10% fetal bovine serum, 30 mM HEPES, 2 mM L-glutamine, and 50 µg/ml gentamicin). NEB-1-P94 or the parental NEB-1 virus were serially diluted in media and 0.2 ml of each dilution inoculated into replicate wells of 96-well plates containing SAM, MA104, or MARC 145 cells. Cultures were incubated for 5 days at 37°C, 3% to 5% CO₂, in a humidified chamber and monitored for cytopathic effects typical of PRRS virus. Titers (50% endpoints) were calculated according to the method of Spearman and Karber. The NEB-1-P94 showed reduced titers or unmeasurable titers on three separate SAM cultures compared to the titers obtained on MA104 and MARC 145 cells (Table 1). This is a phenotypic change compared to the parental strain which showed similar titers on all of the cultures tested. Therefore, impaired growth on swine alveolar macrophages was a selected phenotypic marker for the vaccine strain NEB-1-P94.

Table 1. Comparison of Growth of the NEB-1-P94 Virus on Various Cell Types

Virus	Titer* on MA104	Titer* on MARC 145	Titer* on SAM 7	Titer* on SAM 8	Titer* on SAM 22
NEB-1-P94	5.3	6.1	≤1.2	2.5	<1.2
NEB-1	5.2	6.5	5.2	6.5	6.5

*Titer = log₁₀ TCID₅₀/ml (limit of detection in this assay = 1.2)

EXAMPLE 2

5 Phenotypic Characterization of NEB-1-P94 for Lack of Virulence in Swine

Four gnotobiotic piglets (seven to 10 days of age) from PRRS seronegative sows were inoculated intranasally (3 ml/nare) with NEB-1-P94 master seed virus ($10^{5.3}$ TCID₅₀/ml). Piglets were
10 observed for clinical signs of respiratory disease and the vaccine strain virus was re-isolated from serum at five days post-inoculation. Serum from the first group of pigs was used to intranasally inoculate a second group of gnotobiotic pigs which were monitored in the same way. This process was repeated for a
15 total of five serial backpassages in piglets in order to determine whether the vaccine strain could revert to a virulent state. Vaccine virus was recovered from each successive animal passage, however, respiratory disease was not observed in the gnotobiotic pigs. In addition, virus isolated from the fifth
20 backpassage pigs was intranasally inoculated into one-week-old and three-week-old conventional piglets (approximately $10^{5.3}$ TCID₅₀/ml was administered per piglet). Animals were monitored for 42 days after virus inoculation and no clinical disease signs (i.e. prolonged high fever, respiratory signs, lung lesions)
25 consistent with virulent PRRS infection were found. Therefore, the NEB-1-P94 virus was concluded to be avirulent for induction of respiratory disease in piglets.

Next the NEB-1-P94 virus was examined for its ability to cause
30 reproductive disease. PRRS seronegative sows at 85 days of gestation were inoculated intranasally (3 ml/nare) with the master seed vaccine strain ($10^{4.5}$ TCID₅₀/ml). All sows farrowed at their expected time and 96% of the piglets were born live and healthy. By comparison, uninoculated control sows gave birth to
35 litters where 87% of the piglets were live and healthy. Therefore, the vaccine strain NEB-1-P94 failed to induced reproduction disease typical of a virulent PRRS virus (See Example 4). These data confirmed the avirulent phenotype of the vaccine strain, NEB-1-P94.

EXAMPLE 3

5 Phenotypic Characterization of NEB-1-P94 for Reactivity with PRRS
Virus-Specific Monoclonal Antibodies

MA104 cells infected with parental strain NEB-1 or the vaccine strain NEB-1-P94 virus were examined for reactivity with monoclonal antibodies specific for the PRRS virus by indirect immunofluorescence. Briefly, 96-well plates of confluent MA104 cells were fixed with 80% acetone for 10 minutes at 2 days after infection with each virus. Monolayers were then incubated with SDOW17, VO17, or EP147 monoclonal antibodies. Following washing, monoclonal antibody reactivity with each virus was detected by incubation with fluorescein isothiocyanate conjugated anti-mouse IgG followed by washing and examination for fluorescence by microscopy. Positive fluorescence was noted with all three monoclonal antibodies for the NEB-1 parental strain (Table 2). However, the vaccine strain, NEB-1-P94, had lost reactivity with the SDOW17 monoclonal antibody but tested positive with the other two monoclonal antibodies. These data indicate that the NEB-1-P94 strain had lost expression of epitope recognized by the SDOW17 antibody. The loss of reactivity with this monoclonal antibody most likely represents genetic mutation in the RNA sequence of NEB-1-P94 which resulted in an altered amino acid sequence in the nucleocapsid protein region recognized by SDOW17.

Table 2. Reactivity of Parental and Vaccine Strain PRRS with Specific Monoclonal Antibodies.

	Reactivity with SDOW17	Reactivity with VO17	Reactivity with EP147
NEB-1-P94	-	+	+
NEB-1	+	+	+

EXAMPLE 4

5 Prevention of Reproductive Disease by Vaccination of Sows with
NEB-1-P94

Vaccine was prepared by inoculation of confluent roller bottles
of MA104 cells with NEB-1-P94 (4 passages from the master seed)
at a multiplicity of infection of approximately 1:10 in EMEM
10 containing 10% fetal bovine serum, 2 mM L-glutamine, and 30 µg/ml
gentamicin. Cultures were incubated for three days at 37°C and
then supernatant fluids were harvested by decanting. Virus
fluids were diluted 50% (v/v) with stabilizer (75 g/L tryptone,
30 g/L dextran, 2 g/L gelatin, 100 g/L lactose, 2 g/L sodium
15 glutamate, 1.05 g/L KH₂PO₄, 2.5 g/L K₂HPO₄, 10g/L albumin
fraction V), frozen, and lyophilized. Vaccine was rehydrated
with sterile deionized water and 2 ml (10^{5.1} TCID₅₀/ml)
administered intramuscularly to gilts four to six weeks prior to
breeding.

20

At 85 days of gestation, vaccinated and unvaccinated control
gilts were challenged by intranasal administration of NEB-1 virus
(approximately 10^{6.3} TCID₅₀). Animals were monitored through
seven weeks after farrowing for signs of fetal or neonatal death
25 attributed to PRRS virus. PRRS viremia developed in 11/12 (92%)
of control sows and 100% of their live-born piglets. PRRS
infection during pregnancy resulted in 16% death loss at
parturition (large mummies and stillborn pigs) in the control
group (Table 3) compared to only 6% death loss in vaccinated
30 sows. In addition, vaccination resulted in a 50% reduction in
the incidence of weak and shaky piglets and 94% reduction in
piglets with low birth weights (weighing less than 2 pounds at
birth) when compared to controls. Vaccination prevented
congenital PRRS as evidenced by the absence of PRRS virus in
35 blood or tissues of any piglets from immunized sows and a 55%
prevention of death loss through 7 weeks of age (Table 3) when
compared to controls. The statistically significant prevention
of death loss and virus infection in vaccinated sows and their
offspring clearly demonstrated the efficacy of this vaccine in
40 preventing the reproductive form of PRRS virus-induced disease.

Table 3. Summary of Reproductive Disease Noted After PRRS Virus Challenge of Vaccinated versus Control Sows

Group	Number of Sows (Ave # Pigs/Litter)	% Large Mummies and Stillborn	% Born Live but Died	% Weak and Shaky	% Piglets Weighing <2 lbs.	7 Week Mortality
Vaccinated	21 (10.8)	6%	2%	3%	1%	17%
Controls	12 (10.1)	16%	3%	6%	17%	38%

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EXAMPLE 5

Prevention of Respiratory Disease by Vaccination of Sows with
NEB-1-P94

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Vaccine was prepared by inoculation of confluent roller bottles of MA104 cells with NEB-1-P94 (at 4 passages from the master seed) at a multiplicity of infection of approximately 1:10 in EMEM containing 10% fetal bovine serum, 2 mM L-glutamine, and 30 μ g/ml gentamicin. Cultures were incubated for five days at 37°C and then supernatant fluids were harvested by decanting. Virus fluids were diluted 50% (v/v) with stabilizer (75 g/L tryptone, 30 g/L dextran, 2 g/L gelatin, 100 g/L lactose, 2 g/L sodium glutamate, 1.05 g/L KH_2PO_4 , 2.5 g/L K_2HPO_4 , 10 g/L albumin fraction V), frozen, and lyophilized. Vaccine was rehydrated with sterile deionized water and 1 ml ($10^{4.9}$ TCID₅₀/ml) administered intramuscularly to PRRS seronegative piglets at three weeks of age.

Four weeks following vaccination, piglets were challenged with virulent NEB-1 PRRS virus by intranasal route as described for gilts (Example 4). Piglets were monitored for respiratory disease signs for 14 days after challenge. All unvaccinated control piglets developed clinical signs of respiratory disease compared with 3/40 (8%) of vaccinated animals. Vaccination resulted in a statistically significant reduction in fever, respiratory signs, and clinical illness in vaccinated animals compared to controls (Table 4). This study clearly demonstrated the efficacy of the vaccine in the prevention of respiratory disease caused by PRRS virus in young pigs.

Table 4. Summary of Clinical Disease in Vaccinated and Control Animals Following PRRS Virus Challenge

	Percentage of Animals with Clinical Signs on Indicated Days														
	0	1	2	3	4	5	6	7	8	9	10	11	12	13	14
Respiratory Signs															
Controls	0	0	0	57	53	37	60	40	37	40	47	40	23	30	33
Vaccinates	0	0	0	8	8	0	2	8	0	2	5	5	5	2	0
Fever >104.5°F															
Controls	3	17	43	73	83	73	66	50	37	57	47	37	20	17	NA
Vaccinates	0	14	20	20	14	10	10	20	2	14	10	10	4	4	NA
Clinical Illness															
Controls	0	0	0	23	60	50	60	47	33	50	50	40	33	27	33
Vaccinates	0	0	0	2	0	0	0	2	0	2	2	2	0	0	0

NA = Data not available

Controls n=30

Vaccinates n=40

CLAIMS:

1. A vaccine suitable for use in the protection of swine
5 against porcine reproductive and respiratory syndrome comprising a PRRS virus essentially corresponding to the isolate NEB-1-P94, deposited at the American Type Culture Collection under accession number VR-2525.
- 10 2. The vaccine of Claim 1 which is in a live, killed or modified live (attenuated) form.
3. The vaccine of Claim 2 wherein the viral agent is modified live (attenuated) and is in a liquid, frozen or dessicated form.
- 15 4. The vaccine of Claim 1 containing an amount of $10^{4.0}$ to $10^{9.0}$ TCID₅₀ of the PRRS virus per ml.
5. The vaccine of Claim 1 where the attenuated virus can be
20 distinguished from wild type forms of the virus based on lack of reactivity with monoclonal antibody SDOW17 and lack of efficient growth on swine alveolar macrophage cells.
6. A method of protecting a pig from clinical disease caused by
25 the porcine reproductive and respiratory syndrome virus comprising administering an effective amount of vaccine of Claim 1 to a pig in need of precaution against disease caused by said virus.
- 30 7. A method of claim 6 wherein said vaccine is administered orally, intranasally, intramuscularly, intradermally, intravenously, or subcutaneously.
8. A method of of diagnosing infection with only the vaccine
35 virus of claim 1 versus infection with wild type virus based on isolation of virus from serum or tissue of an infected pig and phenotypic characterization as specified in claim 5.

9. A method of claim 8 wherein diagnosis of infection with vaccine virus or wild type virus comprises by measurement of antibodies in the infected swine that are reactive with the PRRS virus protein epitope recognized by monoclonal antibody SDOW17.
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INTERNATIONAL SEARCH REPORT

Intern al Application No
PCT/US 97/02562

A. CLASSIFICATION OF SUBJECT MATTER IPC 6 A61K39/12 G01N33/569		
According to International Patent Classification (IPC) or to both national classification and IPC		
B. FIELDS SEARCHED Minimum documentation searched (classification system followed by classification symbols) IPC 6 A61K G01N		
Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched		
Electronic data base consulted during the international search (name of data base and, where practical, search terms used)		
C. DOCUMENTS CONSIDERED TO BE RELEVANT		
Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
P,X	WO 97 00696 A (REGENTS OF THE UNIVERSITY OF MINNESOTA) 9 January 1997 see claims 1-8,13-15 ---	1-9
P,X	WO 96 36356 A (BOEHRINGER INGELHEIM CORP.) 21 November 1996 see claims 1-7; example 1 ---	1-9
X	EP 0 676 467 A (AKZO NOBEL N.V.) 11 October 1995 see claims 1-10 ---	1-4,6-9
Y		5
X	GB 2 282 811 A (IBERICA CYANAMID S.A.) 19 April 1995 see claims 1-27 ---	1-4,6-9
Y		5

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Date of the actual completion of the international search 10 June 1997		Date of mailing of the international search report 07.07.97
Name and mailing address of the ISA European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel. (+ 31-70) 340-2040, Tx. 31 651 epo nl, Fax (+ 31-70) 340-3016		Authorized officer Olsen, L

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C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT		
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Y	see claims 10-23	5
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